Nitrogen and Phosphorus Uptake Efficiency and Partitioning of Container-grown Azalea During Spring Growth

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ABSTRACT. The influence of fertilization rate on nitrogen (N) and phosphorus (P) nutrient partitioning and uptake efficiency of young, container-grown azalea (Rhododendron L. ‘Karen’) was determined under controlled greenhouse conditions during Spring 2001 and 2002. In 2001, fertilizer treatments included a factorial combination of two N (25 or 250 mg/week) and three P (0, 5, or 25 mg/week) rates; in 2002, an additional N rate (100 mg/week) was included in the experimental design. Five destructive harvests were performed during each study; plant tissues (root, stem, primary and secondary branches and leaves) from each harvest were analyzed to derive total N and P uptake. Leachates from containers were monitored and analyzed weekly to calculate nitrate (NO$_3$-N), ammonium (NH$_4$-N), and orthophosphate (PO$_4$-P) loss. Fertilization rates of 5 mg P per week in 2001 and rates of 100 mg N per week and 5 mg P per week in 2002 maintained optimal growth compared with the highest fertilization rates (250 mg N and 25 mg P per week) in these studies. Increasing N fertilization rate largely promoted shoot growth, whereas decreasing N and P fertilization rates promoted root growth and increased uptake efficiency. In general, increasing N and P fertilization rates increased nutrient N and P leaching from the pine bark substrate. Reducing excess N and P fertilization to match plant growth requirements of young azalea increases nutrient uptake efficiency and reduces nutrient loss to the environment.
promote root growth or influence root-to-shoot ratio in a wide range of plants (Broschat and Klock-Moore, 2000; Dufault and Schultheis, 1994; Melton and Dufault, 1991; Weston and Zandstra, 1989; Yeager and Wright, 1982).

We hypothesize that current N and P application rates to many ornamental species in container production exceed normal plant N and P growth requirements resulting in low uptake efficiencies and excessive nutrient loss through leaching. A review by Chen et al. (2001) noted that recommended N rates for greenhouse-grown azalea were 2200 kg·ha⁻¹ per year, over 10 times the agronomic rate for corn. This equates to applying 500 mg N per week to a plant in a 7.6-L container over a 40-week growing cycle (given 110,000 × 7.6-L containers per hectare). Borch et al. (1998) noted that levels of P fertilization are orders of magnitude greater than plant requirements; plants grow well at P concentrations 100-fold less than traditional rates used in container plant production (Lin et al., 1996; Lynch et al., 1991) as long as available P concentrations are held constant in the soil solution.

Specifically, we hypothesize that rates of 100 mg N and 5 mg P per plant each week will 1) maintain maximal shoot and root growth rates of young (less than 2 years old) azalea plants, and 2) increase N and P uptake efficiency and reduce N and P leaching (loss) from containers. Azalea was chosen as a model ericaceous ("low nutrient use") woody perennial, because it is widely grown in the nursery and landscape industry throughout the United States. There are few data on nutrient uptake and use efficiency for container-grown azalea. A few studies on other species such as poinsettia (Euphorbia pulcherrima Wild.) (Ku and Hershey, 1992; Rose et al., 1994) or in woody ornamentals (Tyler et al., 1996) have examined nutrient use from the perspective of reducing nutrient concentrations in fertilizer applications, aiming to reduce leaching without impacting plant growth or quality. Research by Sandrock et al. (2005) investigated N nutrition of Weigela florida Bunge ‘Red Prince’ and Euonymus alatus Thumb. ‘Compactus’ based on periodicity of growth and nutrient requirements. Other researchers have studied nutrient uptake and seasonal partitioning physiology of Rhododendron ferrugineum L. in natural ecosystems, contributing much to the knowledge base (Lamaze et al., 2003; Pasche et al., 2002). The objectives of this study were to investigate growth and nutrient uptake of 12- to 18-month-old azalea plants under different combinations of N and P fertility, and recommend fertility rates that optimize growth and nutrient uptake efficiency, yet minimize nutrient leachate potential.

Materials and Methods

**Spring study 2001.** An initial experiment was conducted over a 10-week period from March to May 2001 to investigate the uptake and partitioning of N and P by ‘Karen’ azalea. Five replicates of six treatments in a completely randomized 2 x 3 factorial design provided (growth) limiting and non-limiting rates of N (i.e., 25 and 250 mg/week, respectively) and limiting, sufficient, and high rates of P (0, 5, and 25 mg/week, respectively) to azalea plants. These rates were extrapolated from previous studies on azalea (Ristvey, 2004) and other information from the literature (Chen et al., 2001).

Twelve-month-old azalea liner plants were purchased in Aug. 2000 from a wholesaler for the 2001 experiment. Liner azaleas were given a 20N–4.4P–16.6K completely soluble fertilizer (Peters Professional General Purpose Water Soluble Fertilizer; Scotts Co., Marysville, OH) at 200 mg L⁻¹ N once per week for 6 weeks. Fertilization was discontinued and azalea liners were not fertilized for 8 weeks before onset of this study to reduce the nutrient reserves of the plants and substrate. Daily watering during this period assured a fully leached substrate. Within the greenhouse, seasonal natural ambient light was provided. Temperatures were controlled within a diurnal range of 20 to 28 °C during the experimental period. Liner plants exhibited a limited dormancy (no shoot growth) during part of December and January, although no dormant buds were formed. A few plants exhibited limited flowering during the study with no more than three blooms on a plant. Those flowers were collected as leaf samples during harvests. Azalea liners were transplanted into 7.6-L Classic #2 plastic pots (Nursery Supplies, Chambersburg, PA) filled with a composted pine bark media consisting of equal parts composted pine bark, sphagnum peat, and rice hulls and amended with iron sulfate (0.22 kg·m⁻³) and Micromax (The Scotts Co.) micronutrients (0.68 kg·m⁻³).

Plants received an initial nutrient application and 10 weekly applications (total of 11 applications) of N and P in an otherwise balanced liquid fertilizer solution (Ristvey, 2004) at the rates specified previously for each treatment throughout the 10-week experimental period. The fertilizer was applied to each plant in 250-mL aliquots once per week. Between fertilizations, all plants were deficit irrigated twice per week (i.e., with a zero leaching fraction by using a balance and records of the initial weight at container capacity); in addition, all plants were watered to excess on the day before each fertilization event and the resulting leachate volume recorded with an aliquot preserved and frozen for nutrient analysis. This irrigation was designed to leach the excess N and P and any other accumulating salts remaining in the substrate from the previous week. The first destructive plant harvest was performed before the onset of the study to provide baseline dry weight and nutrient content data. Five further plant harvests were conducted every 2 weeks to provide sequential plant N and P uptake, partitioning, and use efficiency data.

**Plant tissue analysis.** During each harvest, the plants were divided into roots, stems, primary and secondary branches, and leaf tissue. Root tissue was separated from stem tissue and cleaned of all substrate by physical shaking and washing with three successive quick rinses of water. Care was taken in recovering all root tissue by draining all wash water, between rinses, through a #30 (600 μm) sieve. Stem tissue comprised the main trunk of the plant. Primary branches included all woody tissue branching directly from the stem. Secondary branch tissue was denoted as all branches other than primary branch tissue. Tissues were separated at each harvest and fresh weights taken. A 10- to 15-g subsample of each tissue from each replicate plant was freeze-dried using a lyophilizer (Labconco, Kansas City, KS). Any additional tissue was dried in a forced-air ventilated oven at 40 °C for 72 to 96 h. Dry weights were measured when lyophilization or drying was complete. Each lyophilized subsample was milled through a 1.0-mm screen (Foss/Tecator Mill, model 1093; Foss, Höganas, Sweden) taking care to clean the mill thoroughly between each sample. All tissues were analyzed for total carbon (C) and total N concentration using a Carlo-Elba Model CE 2000 CN analyzer (CE Elantech, Lakewood, NJ). Precise analytical sample dry weights were noted for each sample to back-calculate total N and P contents. Tissues from the initial, third, and final harvests...
of each data set were analyzed for total P using an open vessel microwave system (Star System 6; CEM, Raleigh, NC). Approximately 0.5 g of each tissue type (0.2 to 0.3 g for leaf tissue) was placed in each microwave vessel containing 0.5 g of potassium persulfate. Tissues were digested, reconstituted, and analyzed with an Alpkem FS 3000 System NP analyzer (O.I. Analytical, College Station, TX) for total P using the ascobic acid method (Clesceri et al., 1989). Total N and P contents of all tissue samples were back-calculated from all total N and P concentration data and from dry weight and sample weight records during this process.

**Leachates.** Throughout the study, the replicate plants that were designated for the final destructive harvest were placed above catchment saucers to retain leachates. Leachates were collected from these plants once per week, on the same day each week, 1 h after a hand-delivered irrigation of 1 L. Leachate volumes were recorded and sample aliquots taken. Sample aliquots were taken in replicate 25-mL scintillation vials and preserved with 50 µL concentrated sulfuric acid (32 N). The samples were kept frozen until analyzed for NO₃-N, NH₄-N, and orthophosphate (PO₄-P) concentration using the FS3000 analyzer. Nitrate-N was analyzed using the automated cadmium reduction method, NH₄-N by the automated phenate analyzer. Nitrate-N was analyzed using the automated cadmium reduction method, NH₄-N by the automated phenate method, and total P and PO₄-P by the automated ascorbic acid reduction method (Clesceri et al., 1989). Nitrogen and PO₄-P leachate contents were calculated by multiplying the concentration of each ion by the total volume of the water collected each week.

**Substrates.** Residual nutrient contents for each treatment’s substrate were quantified at the end of the study. Subsamples from each replicate within each treatment were combined and one representative sample of ≈1 L in volume was analyzed for each treatment. Substrate analysis was quantified by using the 1:1.5 volume extract method as detailed in Handreck and Black (1994). Each substrate extract was then colorimetrically analyzed using the Alpkem FS 3000 for NH₄-N, NO₃-N, and PO₄-P (ortho-P).

**Initial data and efficiency calculations.** In each study, N and P plant tissue concentrations were multiplied by tissue dry weight to give tissue nutrient content, which normalized differences in nutrient concentrations as a result of growth differences between treatments. The initial harvest (n = 15 plants) was performed before the onset of each study to provide baseline dry weight and N and P content data. The baseline data were then subtracted from each harvest value for each treatment, giving total dry weight and N and P uptake for each treatment every 2 weeks. Because both dry weight and nutrient uptake accumulation values were transformed from dry weight and total nutrient content values using the initial values, both sets of data have the same treatment variances. Uptake data were used to develop the nutrient budgets. The initial harvest at the beginning of the experiment gave a per-plant average of 5.7 g dry weight, 91.6 mg N and 20.9 mg P in the 2001 study, and 7.5 g dry weight, 104.0 mg N and 19.0 mg P in the 2002 study. Plant N and P uptake values were then used to calculate uptake efficiencies. Nitrogen and P uptake efficiency percentages thus only reflect the fraction of applied nutrient accumulated by plants during each 10-week period.

**Spring study 2002.** A repeat experiment, with improvements, was conducted over a similar 10-week period from the end of March through May 2002. Azalea cuttings from mother plants of the original stock were taken in June 2001 and rooted in Oasis Horticubes (Oasis Products, Kent, OH) with Hormodin (OH, Mainland, PA) rooting hormone. After 8 weeks under 4 s mist every 4 min, root cuttings were transferred to 10.2-cm pots containing a 1 peat:1 vermiculite potting substrate. Preexperimental fertilization of azaleas was similar to previous study, but for 14 weeks. Fertilization discontinued 10 weeks before 2002 study initiation. In this study, nine treatments with three replicates arranged in a completely randomized 3 x 3 factorial design provided limiting, sufficient, and high rates of N (i.e., 25, 100, and 250 mg N per week, respectively) combined with limiting, sufficient, and high rates of P (0, 5, and 25 mg P per week, respectively) to azalea plants. Plants received 18 applications of N and P in an otherwise balanced liquid fertilizer solution at the weekly rates specified for each treatment throughout the experimental period. The fertilizer was applied to each plant in 250-mL aliquots twice per week at half rate to minimize any potential salinity effects of the higher rate treatments. Additionally, before the start of the study, an analysis on N and P availability was performed on the substrate. All other methods and procedures were the same as for the first study. Several weeks into the study (at harvest 1), plants began showing signs of a leaf-tip necrosis in all treatments. An immediate foliar and substrate analysis revealed a high concentration of manganese (Mn) in both leaves and substrate. It is possible that the Mn originated in the pine bark used in the substrate. Some sources of fresh bark have been documented to have excessive levels of Mn (Solbraa and Selmer-Olsen, 1981). To counteract the effects of Mn accumulation in plant tissues, sodium silicate was added to each fertilizer solution at a 0.05 M concentration. According to Marschner (1995), silicon tends to prevent Mn accumulation in any one area and promotes the distribution of Mn more evenly throughout plant tissues. The addition of sodium silicate ameliorated the Mn toxicity on all new leaf growth by the third harvest.

**Statistical analysis.** All sample data were analyzed using a factorial analysis of variance using the PROC MIXED routine (SAS Institute, Cary, NC). If treatment interaction was not significant, main effects are reported and discussed. However, if treatment interactions were significant, simple effects (the effect of a variable at a specific level of another variable) are reported and discussed. Pairwise comparisons were done under least significant differences (LSD) criteria at P ≤ 0.05 to increase the power of the test and protect the analysis interpretation from incorrectly accepting the null hypothesis (type II error). The use of LSD increases the power of the test and hence, increases the probability of detecting a real effect.

**Results.**

**Dry weight.** Both studies show that high nutrient application rates do not significantly increase total plant dry weight or shoot dry weight over the moderate N and P rates in this study. Azalea total dry weight results from each final harvest are summarized and compared in Table 1. There was no difference in total dry weight or shoot dry weight between the P at 25 mg/week and P at 5 mg/week treatments combined with N at 250 mg/week (250N:25P and 250N:5P) during either year’s study. In the 2002 study, there were no differences in total and shoot dry weight between the N treatments of 250 mg/week and 100 mg/week regardless of P treatment. In the 2001 study, root dry weight of azaleas given the 25N:0P treatment was significantly greater than the 250N:25P treatment. This result was seen again
Table 1. Average dry weight data and root:shoot ratio in azalea (less than 2 years old) after 10 weeks of treatment (March to May) in 2001 (n = 5) and 2002 (n = 3).*  

<table>
<thead>
<tr>
<th>Treatment (mg/week)</th>
<th>Total dry wt (g)</th>
<th>Shoot dry wt (g)</th>
<th>Root dry wt (g)</th>
<th>Root:shoot (ratio)</th>
</tr>
</thead>
<tbody>
<tr>
<td>250N:25P</td>
<td>25.2 a**</td>
<td>23.4 a</td>
<td>21.9 a</td>
<td>18.7 a</td>
</tr>
<tr>
<td>250N:5P</td>
<td>23.3 ab</td>
<td>22.9 ab</td>
<td>20.1 ab</td>
<td>17.9 a</td>
</tr>
<tr>
<td>250N:0P</td>
<td>19.8 bc</td>
<td>21.6 abc</td>
<td>17.3 bc</td>
<td>16.9 a</td>
</tr>
<tr>
<td>100N:25P</td>
<td>—</td>
<td>22.5 a</td>
<td>—</td>
<td>16.8 a</td>
</tr>
<tr>
<td>100N:5P</td>
<td>—</td>
<td>21.0 abc</td>
<td>—</td>
<td>15.7 ab</td>
</tr>
<tr>
<td>100N:0P</td>
<td>—</td>
<td>23.2 a</td>
<td>—</td>
<td>17.4 a</td>
</tr>
<tr>
<td>25N:25P</td>
<td>16.5 c</td>
<td>17.7 bc</td>
<td>13.3 d</td>
<td>11.6 bc</td>
</tr>
<tr>
<td>25N:5P</td>
<td>18.6 c</td>
<td>16.3 c</td>
<td>14.9 cd</td>
<td>9.9 c</td>
</tr>
<tr>
<td>25N:0P</td>
<td>19.4 bc</td>
<td>17.3 c</td>
<td>15.3 cd</td>
<td>11.5 bc</td>
</tr>
</tbody>
</table>

Significance

N: ***  ****  ****  ****  ****  **  **  —  —
P: NS  NS  NS  NS  NS  NS  NS  —  —
N × P: *  NS  *  NS  *  NS  NS  —  —

*In 2001, azalea were fertilized 11 times over the study period, an initial and 10 weekly applications. In 2002, 18 half-rate applications were made over the study period. Lower case letters indicate significant differences (LSD at P = 0.05) between treatments. Total dry weight refers to total plant dry weight.

Means within columns followed by the same letter are not significantly different (α = 0.05) by LSD.

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treatment compared with all other treatments (Table 2). In the 2002 study, P content was also a function of both N and P rate but with no treatment interaction. In the 2002 study, no differences in total, shoot, or root dry weight among the three treatments, greater P content in the 250N:25P treatment showed luxury consumption and storage with fertilizer containing an N:P ratio of 10:1. Azaleas in the 250N:25P treatment stored excess P mainly in leaf and root tissues. Root P content declined with decreasing P rate within each N treatment although there was little difference in root weight among most treatments. Consequently, the N:P ratio increased directly as a function of decreased P application. Higher N:P ratios were seen in the high N treatment combinations.

**Leaching and Uptake Efficiency.** Differences in N leachate were the result of N treatment alone (Table 3). As N fertilization rate increased, significantly greater amounts of N leached from containers. In the 2002 study, 40 times less N was leached from the N treatments of 25 mg/week, and up to four times less N was leached from the N treatments of 100 mg/week, compared with the N treatments of 250 mg/week (Table 3).

Differences in P leachate were the result of interactive effects between N and P treatment in only the 2001 study. In general, higher P fertilization significantly increased P leachate. Oddly, the 250N:25P treatment averaged significantly less leachate than either the 100N:25P or 25N:25P treatments, despite no obvious differences in growth between the 250N and 100N treatments. Compared with lower nutrient treatment rates, a substantial amount of N and P remained in the substrates of the high N and P treatment rates on completion of both studies (Table 3).

In both studies, N uptake efficiency decreased as N application rate increased and this relationship was more pronounced for P uptake efficiency (Table 3). In the 2002 study, average N uptake efficiencies were significantly greater in azaleas under the N at 25 mg/week than the N at 250 mg/week. Average N uptake efficiencies for azaleas under the N at 100 mg/week were intermediary between the high and low N treatments. The P uptake efficiency of azaleas under the P treatment of 5 mg/week increased threefold over azalea given 25 mg/week of P.

**Discussion**

With regard to the rates used in this study, it should be noted that many growers consider rates of 150 to 200 mg·L⁻¹ N as a “low” N rate, which, if applied as a constant feed (say applying 500 mL fertilizer per day) equates to N rates between 525 and 700 mg/plant per week. This gives a perspective on the rates used in these studies, the highest rate being N at 250 mg/plant per week. Alternatively, many controlled-release fertilizer manufacturers make rate recommendations based on container size. Nitrogen rates of 100 mg/week is considered low for the Custom #2 containers used in this study. For example, the N rate of 100 mg/week is similar to applying 22 g of an 18% nitrogen fertilizer over a 40-week growing season.

The 250 mg/week N rates promoted more pronounced differences in shoot growth compared with root growth of azalea in both studies. More importantly, the “intermediate” N rate of 100 mg/week adequately maintained optimal growth.
Table 2. Average nitrogen and phosphorus content (= concentration × dry weight) in azalea (less than 2 years old) tissues after 10 weeks of treatment (March to May) in 2001 (n = 5) and 2002 (n = 3).

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Total plant N (mg)</th>
<th>Shoot N (mg)</th>
<th>Root N (mg)</th>
<th>Total plant P (mg)</th>
<th>Shoot P (mg)</th>
<th>Root P (mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>250N:25P</td>
<td>523.2 a</td>
<td>428.9 a</td>
<td>455.1 a</td>
<td>332.7 a</td>
<td>68.0 a</td>
<td>96.1 a</td>
</tr>
<tr>
<td>250N:5P</td>
<td>442.9 ab</td>
<td>380.3 ab</td>
<td>383.4 ab</td>
<td>299.2 a</td>
<td>59.6 a</td>
<td>81.1 ab</td>
</tr>
<tr>
<td>250N:0P</td>
<td>401.2 b</td>
<td>417.4 ab</td>
<td>361.6 b</td>
<td>339.8 a</td>
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<td>100N:25P</td>
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<td>—</td>
<td>—</td>
<td>355.3 c</td>
<td>50.3 c</td>
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<td>—</td>
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<td>25N:25P</td>
<td>146.5 c</td>
<td>188.0 c</td>
<td>119.5 c</td>
<td>137.7 b</td>
<td>27.0 a</td>
<td>74.1 abc</td>
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<tr>
<td>25N:5P</td>
<td>152.4 ab</td>
<td>186.8 c</td>
<td>124.8 ab</td>
<td>127.0 b</td>
<td>51.7 b</td>
<td>59.8 ab</td>
</tr>
<tr>
<td>25N:0P</td>
<td>216.4 c</td>
<td>177.9 c</td>
<td>182.6 c</td>
<td>126.6 b</td>
<td>8.1 a</td>
<td>10.8 ab</td>
</tr>
</tbody>
</table>

Significance

<table>
<thead>
<tr>
<th>N</th>
<th>P</th>
<th>N × P</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
</tbody>
</table>

The response of root growth was quite different and was contradictory to the conventional view that high P fertilization increases root growth (Harris, 1992). The greatest root:shoot ratios were with the 25 mg/week (low) N treatments because shoot growth was limited by available N. In the first study, the greatest root dry weight was seen in the 25N:0P treatment; whereas root growth was reduced without P in the 250N:0P treatment, this was not significantly different from any high N treatment or the 25N:25P treatment, again perhaps as a result of native P in the substrate. Thus, in azalea (a low nutrient use species), root weight is not improved with high P fertilization but increases under conditions of low nutrient concentration as has noted by other authors for other plant species (Borch et al., 1998; Hansen and Lynch, 1998; Lynch et al., 1991; Yeager and Wright, 1981; Zhang et al., 2002). In the second study, P treatments had no effect on root growth, most likely attributable to the presence of adequate amounts of plant-available P in the substrate throughout the study. Sufficient rates of both N and P (100N:5P) sustained root growth that was no different from the N treatments of 25 mg/week. A comparatively larger average root weight was found with low N treatments. Fertilization recommendations commonly focus on promoting maximum shoot growth in plants. By using more moderate N rates, more optimal root/shoot ratios can be maintained, which may improve nutrient uptake efficiencies while in production and promote greater postplanting survival rate in the landscape.

Although supplying the azalea with high N and P rates increased plant nutrient contents, greater quantities of residual substrate N and P were available for loss through leaching and perhaps other loss mechanisms. There was a surprisingly large quantity of residual N and P left in substrates at the end of the study, especially from the high N and P treatments (Table 3). Although Marconi and Nelson (1984) concluded that soilless mixes had low P adsorption capacity and PO₄³⁻ ions could leach easily from these types of substrate, they also found in some cases that P did not leach out in expected quantities. Marconi and Nelson (1984) found that irrigation water did not achieve 100% displacement throughout the substrate and that the applied water channeled through macropores, leaching out only portions of applied P. It is reasonable to suspect that this may have occurred during this study with both N and P. Additionally, azalea roots did not completely explore the container volume during the study, leaving areas within the container untouched by root activity.

In both studies, average P leachate was greatest in the 25N:25P treatment, most likely as a result of the N growth limitation; consequently, the applied P was not fully used by the plant. In the 2001 study, there was some P leachate from the zero P treatments, perhaps as a result of root turnover or the release of native P by the pine bark substrate. An even greater amount of available P was found in the second study’s substrate in amounts enough to sustain growth without P fertilization. This may have been because substrate was not well composted (as evidenced by the manganese problem). In fact, the second highest average dry weight was sampled from the 100N:0P treatment. Despite many publications pointing to the contrary (Gutschick and Kay, 1995; Hansen and Lynch, 1998, Lynch et al., 1991), the belief that it is necessary to apply more than minimal quantities of P to promote root development seems to
be common in the nursery industry. Fertilizers containing near equimolar ratios of N and P claim to boost root growth. The results presented here add to the growing literature disputing these assertions. Because no differences in growth were noted with either P treatment of 25 mg/week or 5 mg/week, optimal P rates should be between 0 and 5 mg/week for young azalea.

Nutrient uptake efficiency was affected primarily by the amount of nutrient applied. The plants under the highest nutrient (250 mg N or 25 mg P) regimes only used between 11% and 16% of the total N and P applied in both studies. Plant uptake efficiencies for N and P were two to four times greater with lower rates of both N and P, indicating that fertilization rates were in excess of immediate, but not necessarily long-term, plant requirements. Interestingly, Sandrock et al. (2005) suggest that uptake efficiencies for lower nutrient rates may be overestimated as a result of available nutrients in the soilless potting substrate. Available N in the soilless potting substrate at initiation of the 2002 study was no more than 8 mg per plant container and when added to the budget had a negligible effect on N uptake efficiencies for all rates. Initial soluble P was ≈29 mg/container. Consequently, recalculating P uptake efficiencies in Table 3 by the addition of this 29 mg available P would have lowered efficiencies by less than 1.5% in the high P treatments of 25 mg/week. Phosphorus uptake efficiencies would have dropped to ≈20% for azaleas given the sufficient P rate of 5 mg/week compared with the calculated range of 40.1% to 49.2% (Table 3). Interestingly, the uptake efficiencies from the no-P rates would have been between 42% and 59%. However, given the variability of readily available nutrients in various soilless potting media, it would not be advisable to base nutrient fertility on potential nutrient availability, because pine bark substrates are extremely variable.

The results of Sandrock et al. (2005) suggest that bound N may be mineralized in soilless potting media, which may lead to

![Fig. 2. Average (n = 3) nitrogen partitioning (cumulative) of azalea root, stem, primary and secondary branch and leaves given (A) 250N:5P, (B) 100N:5P, and (C) 25N:5P mg/week from 25 Mar. to 20 May 2002. The top partitioning line represents total N content. Areas between lines represent N partitioned by tissue. Error bars about the mean represent 1 se.](image1)

![Fig. 3. Average (n = 3) phosphorus partitioning (cumulative) of azalea root, stem, primary and secondary branch and leaves given (A) 250N:25P, (B) 250N:5P, and (C) 250N:0P mg/week from 25 Mar. to 20 May 2002. Results shown are from day 0, day 42, and day 70 harvests. The top partitioning line represents total P content. Areas between lines represent P partitioned by tissue. Error bars about the mean represent 1 se.](image2)
### Table 3. Nutrient uptake and nutrient budget for azalea (less than 2 years old) after 10 weeks of treatment (March to May) in 2001 (n = 5) and 2002 (n = 3).  

<table>
<thead>
<tr>
<th>Treatment (mg/week)</th>
<th>Plant N uptake (mg)</th>
<th>N Leachate (mg)</th>
<th>Substrate N (mg)</th>
<th>N uptake efficiency (%)</th>
<th>Plant P uptake (mg)</th>
<th>P Leachate (mg)</th>
<th>Substrate P (mg)</th>
<th>P uptake efficiency (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>250N:25P</td>
<td>431.6 a</td>
<td>324.9 a</td>
<td>84.2 a</td>
<td>117.1 b</td>
<td>695.6</td>
<td>762.6</td>
<td>15.8 bc</td>
<td>14.4 bc</td>
</tr>
<tr>
<td>250N:5P</td>
<td>351.4 ab</td>
<td>276.3 ab</td>
<td>99.2 a</td>
<td>152.3 a</td>
<td>588.3</td>
<td>620.3</td>
<td>12.8 c</td>
<td>12.3 c</td>
</tr>
<tr>
<td>250N:0P</td>
<td>309.6 b</td>
<td>313.4 ab</td>
<td>106.7 a</td>
<td>147.6 a</td>
<td>670.6</td>
<td>897.1</td>
<td>11.1 c</td>
<td>13.9 bc</td>
</tr>
<tr>
<td>100N:25P</td>
<td>54.9 c</td>
<td>84.0 c</td>
<td>9.7 b</td>
<td>13.6 a</td>
<td>178.0</td>
<td>26.9 abc</td>
<td>17.6 a</td>
<td>16.4</td>
</tr>
<tr>
<td>250N:0P</td>
<td>80.2 c</td>
<td>73.9 c</td>
<td>12.0 b</td>
<td>17.3 ab</td>
<td>232.0</td>
<td>30.7 abc</td>
<td>24.3 ab</td>
<td>15.3</td>
</tr>
</tbody>
</table>

Significance

- **Means within columns followed by the same letter are not significantly different (LSD at \(\alpha = 0.05\)).**
- Means within columns followed by the same letter are not significantly different (LSD at \(\alpha = 0.05\) by LSD).
- **NS** = Not significant.

### Literature Cited


